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Delphinid brain development from neonate to adulthood with comparisons to other cetaceans and artiodactyls

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ABSTRACT

Why do neonatal and adult delphinids have much larger brains than artiodactyls when they have common ancestors? We explore relationships between neonatal brain size, gestation duration, maternal body mass, and body growth. Cetacean brains grow fast in the womb and longer gestation results in a larger brain. Allometry shows that the larger the mother's body mass, the larger the neonatal brain. After birth, delphinid bodies grow much faster than brains, and the index of encephalization decreases even as brains grow beyond maturity. Delphinids' larger brain growth during life at sea may be explained by at least three differences from artiodactyls' life on land. First, the sea offers high calorie prey to support growth of a large brain. Second, life in water offers relief from gravity, allowing for a large head to contain a large brain. Third, sound in water may pass through an immersed body. This allows sounds from the water to reach the fetus, driving early development of delphinoid auditory brain parts. As an example of this, the dolphin ear bone is very large at birth. Furthermore, the auditory nervous system appears mature well before birth. Compared with artiodactyls, these three differences likely result in a larger delphinid brain.

Key words: dolphin, porpoise, whale, cetacean, brain size, brain growth, neonate, gestation.

Around fifty million years ago, some ancestors of artiodactyls began to leave the land for a life in water (Gingerich *et al.* 2001, Berta *et al.* 2005) and became the animals we know as cetaceans (whales, dolphins, and porpoises). Now these animals belong to a larger order of mammals, the Cetartiodactyla (cetaceans and artiodactyls). In leaving the land, cetaceans were freed from some gravitational constraints that might limit their size. They also gained access to a rich, high calorie food supply that enabled many of them to grow very large. We have long known that cetaceans have large brains but data on growth of cetacean brains across the lifespan are extremely rare.

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Brain size of terrestrial mammals has long been studied. These data offer insights into the life history, metabolism, function, ecology, and evolution of these animals (Harvey and Krebs 1990, Barton and Capellini 2011, Boddy et al. 2012, Isler and van Schaik 2012). Although body growth curves have been presented for some cetaceans involved in commercial whaling or fishery bycatch (Laws 1959, Lockyer 2007 and references therein), very little data are available on growth of the brain in most cetacean species. Often, brain measures of individuals are presented without companion information on maturity. Although scarce, ontogenetic data on brain mass relative to body mass have been presented for some cetacean species (Perrin et al. 1977, Miyazaki et al. 1981, McLellan et al. 2002, Mallette et al. 2016). With the exception of these aforementioned studies, some species have been represented by inaccurately measured data in the literature.

For instance, an erroneous mass from an alcohol-dehydrated blue whale (*Balaenoptera musculus*) brain represented the species in the literature for over 100 yr (Ridgway and Van Alstyne 2017). Another problem arose from the use of endocasts as a proxy for brain mass (Jerison 1973; Marino *et al.* 2000, 2004; Montgomery *et al.* 2013). The cranial vault of cetaceans, especially in larger whales, contains large vascular networks and tentorium cerebelli that may take up to 65% of the cranial vault volume (see table 3 in Ridgway *et al.* 2017). Without taking this nonbrain tissue into consideration, using endocasts alone may lead to overestimation of brain size. Scarcity of cetacean brain mass data in the literature, particularly from the mysticetes, makes erroneous data that much more detrimental to the progress that is being made to understand cetacean brain-body relations.

Furthermore, much of the current data on cetacean brains come from both mature and immature animals, but with little data on neonatal brains. The need for data on the size of the brain at birth and its growth to maturity is often mentioned in the literature (Sacher and Staffeldt 1974, Marino 1997, Barton and Capellini 2011, Isler and van Schaik 2012). With an extensive data set collected over 50 yr, we seek to highlight important differences in the brains of cetaceans based on maturation.

Sacher and Staffeldt (1974) demonstrated a positive allometric relationship between gestation duration and adult brain mass in 91 species of homeothermic placental mammals. Weisbecker and Goswami (2010) also found evidence of this pattern in another study of precocial placental mammals and formulated the "Neonatal Maturity Hypothesis" based on the pattern. The literature is conflicted as to whether there is evidence of this pattern in cetaceans. Perrin *et al.* (1977) found support for this pattern in stenellids, but Marino (1997) found no such evidence. In the years since these publications, various authors have provided more precise data on cetacean gestation time (*e.g.*, Robeck *et al.* 2004, 2009; O'Brien and Robeck 2012) and on neonatal brain size (Ridgway and Hanson 2014). The current study provides a much larger data set of cetaceans, with specimens of various developmental stages, to address whether the Neonatal Maturity Hypothesis is supported for cetaceans.

In addition to faulty data and failing to account for differences due to maturity, cetaceans have been mistakenly generalized in some literature, suggesting that all adult cetaceans have large brains relative to their bodies. However, Ridgway *et al.* (2017) demonstrated great variation in cetacean brain size relative to their body size. In addition to variations in gross brain morphology, brain parts also vary by taxa. For example, delphinids have the largest cerebellums relative to body mass and to body length of all cetaceans.

This study presents allometric relationships between cetacean neonatal brain size and body growth for the first time. The data reported in this study come from cetaceans with known body mass, body length, and maturity. Limited results have been previously published by Ridgway et al. (1966), Ridgway (1981, 1986, 1990), Tarpley and Ridgway (1994), Ridgway and Tarpley (1996), Marino et al. (2000), Hanson et al. (2013), Ridgway and Hanson (2014), and Ridgway et al. (2017). The goal of the present study was to elucidate delphinid brain relationships among the genera and species: (1) neonatal brain mass vs. mother's body mass, (2) index of encephalization for adult vs. neonate brains, and (3) brain growth trends across their lifespan. We also wanted to compare neonatal brain size against gestation duration to test the hypothesis that species with longer gestations produce larger brains. More broadly, we wanted to compare brain mass and growth of Delphinidae with the small amount of data available for other Cetacea and Artiodactyla.

MATERIALS AND METHODS

The species included in our data set are listed by taxonomic family in Table S1. Our sample of adult, subadult, and neonatal cetaceans represented both odontocetes (10 families, 29 genera, 42 species) and mysticetes (3 families, 5 genera, 9 species). For each species, Table S1 also displays the number of specimens from each of the three age groups. Data on body length permitted maturity estimation (Perrin *et al.* 1984), allowing us to categorize animal maturity with respect to body length. Neonates were classified as animals that were 60 d or younger. Animals were classified as adults if they exceeded mature body length (see Table S1), and animals were classified as subadults if they did not exceed mature length but were older than 60 d.

Brain masses presented in this study were measured directly by the authors, extracted from the literature, or shared by colleagues. The origin of each data point is noted in Table S1. Data sources are listed as "author," "other," and "lit." The authors in this study directly measured the "author" data points. A recent open access paper covering only adult cetacean brains presents more detailed information on methods for brain measures (Ridgway et al. 2017). "Lit" data points were extracted from the literature, and "other" data points were previously unpublished and came from personal correspondence, stranding events, etc. Observed gestation durations came from the literature (Hayssen et al. 1993, Robeck et al. 2004, O'Brien and Robeck 2012). All data points came from individual animals. Neonatal data presented here are mainly from brains extracted after fishery bycatch and brains extracted from animals that died of natural causes in human care or stranded on beaches.

Cetacean brains appear to contain slightly more white matter (DeGraaf 1967), and white matter is heavier than gray matter (Gompertz 1902). Thus, specific gravity of cetacean brains is slightly higher than the 1.036 generally stated for the human brain (Gompertz 1902). Specific gravity was determined in a subset of cetacean brains by immersion in water (specific gravity 1.0). The amount of water displaced is equal to brain volume. The mass of the brain in air minus the immersed mass is divided by the mass in air to yield the specific gravity of our cetacean brains, which was 1.04. Most brains were not weighed while immersed, thus brain volume was calculated by dividing brain mass by 1.04, the mean specific gravity calculated for cetacean brains. For our regressions, we have used both brain mass and brain volume.

All linear regressions and resulting linear equations were generated in XLSTAT (2016.3, Statistical Innovations Inc., Boston, MA). Before completing the regression analyses, brain mass, body mass, and gestation duration were log-transformed (with base 10) to derive a linear equation that describes an allometric (biological scaling)

relationship between two of the variables. In addition to testing the allometric relationship between gestation duration and mother's body mass with brain size, regressions were also used to present allometric relationships between brain size and maturity. We used the formula of Jerison (1973) to compare the index of encephalization (EQ) across maturity in six odontocete species: $EQ_i = \frac{E_i}{0.12P_i^{2/3}}$ (EQ_i = index of encephalization for species *i*; E_i = brain mass in *g* for species *i*; P_i = body mass in *g* for species *i*). Jerison's EQ is a number that represents how much larger or smaller a brain is relative to the expected brain size based on body size.

RESULTS

Factors Affecting Neonatal Brain Size

Our data show that, across four species of delphinids, neonatal brain mass scales to the 0.51 power of maternal body mass (Fig. 1); maternal body mass spanned a wide range for these four species, from 60 kg in *C. commersonii* to over 3,000 kg in *O. orca.* We also used our data to compare adult and neonatal EQ. The extent to which neonatal and adult animals of 15 cetacean species differ in EQ is displayed in Table 1. We also compared neonatal brain volumes with gestation durations by species. There is a linear relationship and strong positive correlation between neonatal delphinids' brain volumes and gestation duration (in days) (Fig. 2). Gestation duration scales to the 0.23 power of brain volume.

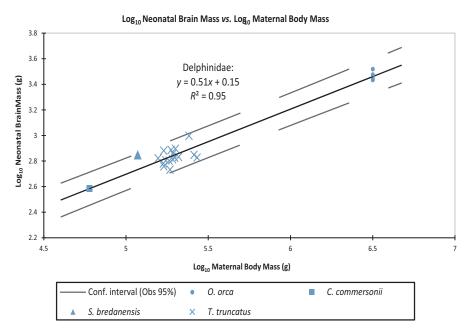


Figure 1. There is a strong, positive correlation between maternal body mass and neonatal brain mass in these four delphinid species; neonatal brain mass scales to the 0.51 power of maternal body mass.

Table 1. Comp	parison of adult and	neonate in	dex of encepha	lization (E	Q) for 1	5 cetacean
(1 mysticete, 14	odontocete) species	s. EQs were	e derived from	brain an	d body	masses in
Table S1.						

Family	Species	Adult EQ	Neonate EQ
Balaenopteridae	B. physalus	0.495	2.297
Delphinidae	C. commersonii	5.149	7.234
*	D. delphis	3.962	7.801
	G. griseus	4.055	5.807
	L. acutus	3.805	6.632
	L. obliquidens	4.635	8.315
	O. orca	2.425	8.317
	S. bredanensis	5.633	8.065
	T. truncatus	3.972	8.328
Physeteridae	P. macrocephalus	0.681	4.402
Kogiidae	K. breviceps	1.703	4.767
Pontoporiidae	P. blainvillei	1.930	2.571
Monodontidae	D. leucas	2.643	5.764
Phocoenidae	P. phocoena	2.837	4.406
	P. dalli	2.909	3.275

Brain Growth from Neonate to Adult

We compared brain mass and maturity of six different odontocete species (four from family Delphinidae, one from Kogiidae, and one from Phocoenidae) for which we had data for various-sized animals from neonates to fully mature adults, with the exception of data from *S. coeruleoalba* that ranged from young subadult to adult age. (Fig. 3). *S. longirostris*, *K. breviceps*, *O. orca*, *T. truncatus*, *S. coeruleoalba*, and *P. dalli* all appear to have brain growth beyond sexual maturity. We also investigated EQ as a function of maturity across six odontocetes, for which we had robust data (Fig. 4), and found that EQ appears to decrease with increasing body length in *O. orca*, *T. truncatus*, *S. coeruleoalba*, *P. macrocephalus*, *P. dalli*, and *K. breviceps*.

Comparison with Terrestrial Relatives

We compared mature body and brain mass between seven terrestrial cetartiodactyls (and an African elephant) with eight delphinids of similar body size. These comparisons are presented in Table 2. The cetacean adults have brains that are approximately 1.5 to almost 10 times larger compared to their terrestrial counterparts of similar body mass. The neonatal comparisons are also quite extreme; as neonatal cetacean brains are about two to three times larger than their terrestrial counterparts.

DISCUSSION

Factors Affecting Neonatal Brain Size

Sacher and Staffeldt (1974) derived a formula relating neonatal brain mass to gestation duration in terrestrial mammals: $\log G = 0.274 \log E_n + 0.144 \log A_e + 0.173 \log N + 1.853$ (G = gestation time in days; $E_n = \text{neonatal brain mass in grams}$; $A_e = \text{ratio of neonatal to adult brain mass}$; N = litter size, 1 in cetaceans). Perrin *et al.* (1977) employed this relationship to predict gestation time in delphinids of the

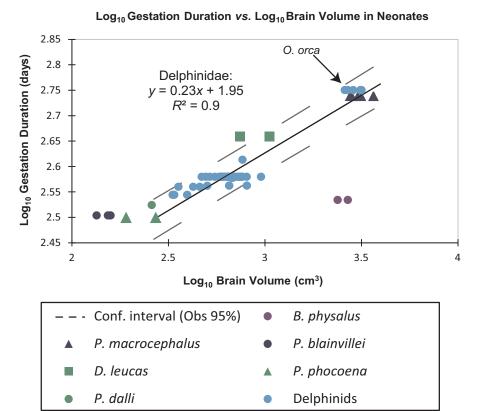


Figure 2. Linear relationship between neonate brain volume and gestation duration (in days). The regression includes only delphinids. Other species were plotted but not included in the regression. The species *O. orca* is indicated by a black arrow. There is a strong, positive correlation between neonatal delphinid brain volume and gestation duration; gestation duration scales to the 0.23 power of neonatal brain volume.

genus *Stenella*. We still do not have data on conception dates and parturition dates (observed gestation times) on that genus for comparison with the prediction from brain mass. Few neonatal cetacean brain masses or volumes have been published in the literature (Perrin *et al.* 1977, see fig. 4B in Mallette *et al.* 2016). It appears that our neonatal brain mass data for *T. truncatus* and for *Stenella* spp. are very similar to the values measured by Mallette *et al.* (2016) and Perrin *et al.* (1977).

Table 3 expands upon the findings of Perrin *et al.* (1977), displaying gestation duration and neonatal and adult brain mass for 17 cetacean species from our data set. For most of the family Delphinidae, we observe a close fit of our data to the Sacher and Staffeldt (1974) formula. The *Phocoena* and *Phocoenoides* data include only three neonates and those from *Delphinus* also represent just three neonates. More data from these species are required to determine if they may better fit the formula. The killer whale has the longest gestation period among the delphinids (and also among all cetaceans), and its observed gestation period and brain mass also fit well with the predicted duration of the Sacher and Staffeldt formula. Our data from delphinids, such

as *T. truncatus* and *O. orca*, demonstrate a very good fit to the formula of Sacher and Staffeldt (1974), as their observed (actual) gestation durations are close to those predicted by the formula (Table 3). Based on known conception dates, mean gestation for *T. truncatus* is 376 d (O'Brien and Robeck 2012), and mean gestation for *O. orca* is 553 d (Robeck *et al.* 2004). The Sacher and Staffeldt formula predicts 377 d for *T. truncatus* and 566 d for *O. orca*.

Unlike delphinid gestation duration, observed mysticete gestation duration does not fit the formula (Table 3). Gestation has been tracked on a large number of mysticetes during whaling activities of the 19th and early 20th centuries. It is likely that gestation estimates from mysticete whales are reasonably accurate. Mysticete whales have relatively short periods of gestation compared to the larger odontocetes, *Physeter* and *Orcinus*, considering their body size and brain size (Table 3). In addition, mysticete whales have much shorter periods of lactation compared to some of the odontocetes (Brodie 1969). Mysticetes are filter feeders, engulfing large amounts of densely spaced prey items (Williams 2006). In contrast, the larger delphinids hunt for individual, more elusive prey. To learn a hunting strategy for individual prey items most likely requires a longer period of learning, and thus a longer period of nursing (Brodie 1969).

Marino (1997) found no relation between EQ and gestation time among 15 odon-tocetes. However, with more precise data on cetacean gestation duration (e.g., Robeck et al. 2004, 2009; O'Brien and Robeck 2012) and our data on neonatal brain mass, we found a significant positive relationship between brain mass and gestation duration across some odontocete species (O. orca, S. bredanensis. C. commersoni, L. obliquidens, T. truncatus, and P. macrocephalus). Therefore, our data support the Neonatal Maturity Hypothesis (Weisbecker and Goswami 2010) for delphinids.

In two delphinid species, *O. orca* and *T. truncatus*, it is obvious that brain growth rate is much higher *in utero* compared to postnatal brain growth rate. For example, brains of *O. orca* require 10 yr or more to double their neonatal brain mass; *T. truncatus* brains may require 8–10 yr to reach full size (Fig. 3). Cranium diameter is likely to be a good predictor of brain growth *in utero*. Each year, numerous pregnant *T. truncatus* are examined with ultrasound. This method has been used to measure the biparietal diameter of the *T. truncatus* fetus at various points of development during gestation (Lacave *et al.* 2004). For this cetacean, it should be possible to develop an accurate profile of brain growth *in utero*.

Figure 5 displays a well-developed neonatal killer whale brain compared to an adult killer whale brain on the same scale. It is interesting to compare the long killer whale gestation with the much shorter gestation of the fin whale (*B. physalus*) (Fig. 2). It appears that the fin whale neonate grows a brain that is only slightly smaller than the killer whale neonate in a much shorter period of time. More data on neonatal brain size in mysticete whales is needed. In our data set, brain mass values for neonatal fin whales are comparable to brain mass values for adult bowhead whales (*B. mysticetus*). Fin whales mature at 5–8 yr (Laws 1959), while bowhead whales do not mature until 12–20 yr of age (Atkinson and Yoshioka 2007); bowhead whales are suggested to have very long life spans, perhaps over 100 yr (George *et al.* 1999, Charvet and Finlay 2012). Bowhead whales also have longer gestation durations compared to fin whales (*ca.* 14.5 mo *vs.* 11–12 mo) (Reese *et al.* 2001). *B. mysticetus* and other right whales have, relative to body size, the smallest brains of all cetaceans (Ridgway *et al.* 2017). More data on neonatal brain size in mysticetes may help us to determine

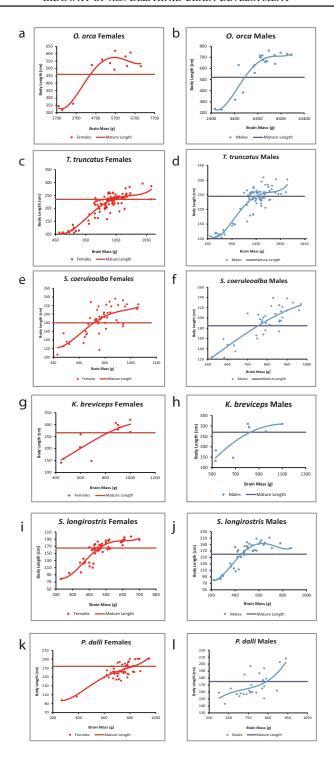


Figure 3. Brain mass relative to maturity (assessed by body length) in six different species. The horizontal line in each species plot represents the length at maturity. Female killer whales (a) (0. orca) reach sexual maturity at about 460 cm body length and as young as 8 yr of age (Dahlheim and Heyning 1999), while male killer whales (b) reach sexual maturity at about 520 cm length when they are around 15 yr of age (Dahlheim and Heyning 1999). Female Common bottlenose dolphins (c) (T. truncatus) reach sexual maturity at a length of 235 cm and at an average age of 8-9 yr (Wells and Scott 1999), and males (d) reach sexual maturity at a length of about 245 cm and an approximate age of 10 yr (Wells and Scott 1999). Female striped dolphins (e) (S. coeruleoalba) reach sexual maturity at 180 cm and about 7 yr of age (Perrin et al. 1994); males (f) reach sexual maturity at about 185 cm and about 11 yr of age (Perrin et al. 1994). Female pygmy sperm whales (g) (K. breviceps) reach sexual maturity at about 266 cm body length (Caldwell and Caldwell 1989), and males (h) reach sexual maturity at about 270 cm length (Caldwell and Caldwell 1989). Female spinner dolphins (i) (S. longirostris) reach sexual maturity at a length of 165 cm and at an average age of 4-7 yr (Perrin and Gilpatrick 1994) while males of this species (j) attain sexual maturity at a length of about 160 cm and an approximate age of 7-10 yr (Perrin and Gilpatrick 1994). Lastly, female Dall's porpoises (k) (P. dalli) reach sexual maturity at 174 cm and about 5 yr of age (Houck and Jefferson 1999), and males (l) reach sexual maturity at about 175 cm and about 5 yr of age (Houck and Jefferson 1999).

if other balaenopterids are born with especially large brains near the size of adult balaenid brains or if the fin whale is an exception.

Brain Growth from Neonate to Adult

An extensive study of over 20,000 fresh human brains, ranging from newborn to over 85 yr of age, revealed that the largest increase in brain mass (for both sexes) occurred during the first 3 yr of life (Dekaban *et al.* 1978). Brain mass increases more than twofold in the first year of life for humans. The neonatal human brain is about 20%–30% of the adult volume (Passingham 1975, Dekaban *et al.* 1978, Dobbing and Sands 1979) compared to about 40% or even 50% or more in many cetaceans (Table 3). Chimpanzee neonates also have brains of about 40% of the adult volume (DeSilva and Lesnik 2006).

We compared brain mass and maturity of four species of delphinids, one phocoenid, and one member of the family Kogiidae (*K. breviceps*) for which we had data from neonates to fully mature animals (Fig. 3). There is considerable evidence among terrestrial mammals that larger brains take longer to grow both in the womb and after birth (Boddy *et al.* 2012). This is also the case for cetaceans we studied. Killer whales, with the largest of the delphinid brains and the longest gestation period of all cetaceans, appear to have continued brain growth past sexual maturity (Fig. 3). Thus, the periods of brain growth in these animals could be as long as 20 yr. Long periods of postnatal growth might support development of social relationships for animals living in stable groups. Connor (2007) presented a case for the involvement of complex social relationships in development of large brains.

Our fresh brain mass values show that EQ in *T. truncatus* is around 8 at birth and decreases as length increases to maturity around an EQ of 4 (Fig. 5). In *O. orca*, EQ is similar to *T. truncatus* at birth but decreases as body length increases to maturity around an EQ of 2. In *K. breviceps*, the pygmy sperm whale, EQ decreases from about 5 to about 1.5, while in its giant relative *P. macrocephalus*, the sperm whale, EQ

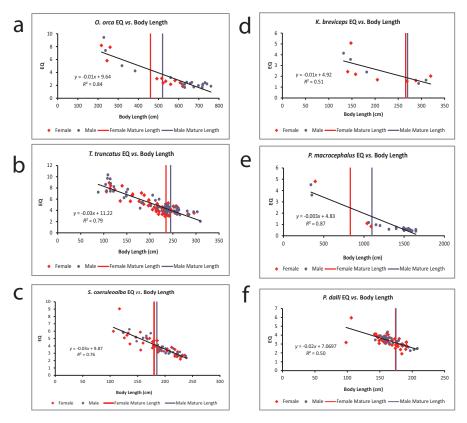


Figure 4. Encephalization quotient (EQ) and body lengths. Body lengths are used as a general indicator for maturity of these animals. Three delphinids (Orcinus orca, Tursiops truncatus, and Stenella coeruleoalba) are compared with EQ and body length against two members of Physeteroidea (Kogia breviceps and Physeter macrocephalus) and one member of Phocoenidae (Phocoenides dalli). In each case, EQ declines as the animal grows toward a mature body length and perhaps beyond. EQ was measured directly from brain masses, except for a few of the larger O. orca for which brain mass was calculated from endocranial volume. Body mass varies considerably in mature animals. As a result, EQ in mature T. truncatus varies from around 3 to 5 and in O. orca from about 1.5 to 3. One outlier EQ value of 2 from a male T. truncatus was from an overweight animal.

decreases from around 4.5 to about 0.5 (Fig. 4). It appears that EQ changes rather dramatically over the course of development in these cetacean species.

Comparison to Terrestrial Relatives

As seen in Table 2, the adult delphinids have brains up to ten times as large as terrestrial cetartiodactyls of similar body size. Delphinid neonates have brains that are about two to three times the size of terrestrial cetartiodactyl neonates of similar body mass. Thus, even with similar sized bodies and, in some cases, shorter gestation

Table 2. Comparison of seven terrestrial cetartiodactyls (and the African elephant) with eight aquatic cetartiodactyls on brain and body mass for neonates and adults. ABoM = adult body mass; ABrM = adult brain mass; NBoM = neonatal body mass; NBrM = neonatal brain mass. All brain and body mass data for the aquatic species come from Table S1.

		ABoM	Ą	\sim	NBrM	Aquatic		Ţ	¥		NBrM
Terrestrial species Commo	Common name	(kg)	(g)	(kg)	(g)	species	Соттоп пате	(kg)	(g)	(kg)	(g)
D. dorcas phillipsi	Blesbok antelope	e^{09}	155^a			— D. delphis	Common dolphin	89	715	11	430
S. scrofa Wild boar	Wild boar	149^{b}	$133^{\rm b}$			L. acutus	Atlantic white-sided	156	1,285	28	733
							dolphin				
T. strepsiceros	Greater kudu	218^{a}	307^{a}			T. truncatus	Bottlenose dolphin	190	1,550	18	685
G. camelopardalis	Giraffe	470^{c}	537°	150^{c}	428^{c}	G. griseus	Risso's dolphin	301	2,132	85	962
C. bactrianus	Bactrian camel	594^{d}	$518^{\rm d}$			G. macrorbynchus	Short-finned pilot	654	2,679		
							whale				
B. taurus	Cow	598°	$492^{\rm e}$	25 ⁸	199^{8}	D. leucas	Beluga	999	2,087	20	938
H. amphibius	Hippopotamus	$1,351^{f}$	720^{f}	40^{8}	195^{8}	G. melas	Long-finned pilot	1,369	3,499		
					,		whale				
L. africana	African elephant	$5,000^{a}$	$4,619^{a}$		$1,724^{\rm h}$	O. orca	Killer whale	3,723	6,642	171	3,006

^aHerculano-Houzel (2015).

^bMinervini et al. (2016).

Graic et al. (2017). ^dXie *et al.* (2011).

Ballarin et al. (2016).

⁸Sacher and Staffeldt (1974). ^hShoshani *et al.* (2006). ^fSilva and Downing (1995).

Table 3. Gestation and brain size. The predicted gestation period was derived by applying the Sacher and Staffeldt formula and using our brain mass data. Sheep (O. aries), cows (B. taurus), giraffes (G. camelopardalis), and hippopotamuses (H. amphibius) were included in the table to compare cetaceans to other members of the Cetartiodactyla taxonomic order. Humans (H. sapiens) were also included for comparison. Cetaceans appear to have similar neonatal/adult brain mass ratios compared to other animals of the Cetartiodactyla order. Sources for the published gestation durations and cetacean brain masses can be found in Table S1.

Taxonomic family Genus species	Neonatal brain mass (g)	Adult brain mass (g)	Neonate/ adult (%)	Published gestation (days)	Predicted gestation (days)
Delphinidae				<u> </u>	
C. commersonii	370	783	47.3	334	324
D. delphis	430	715	60.2	363	359
G. griseus	796	2,132	37.3	410	386
L. acutus	733	1,285	57	365	401
L. obliquidens	523	1,198	43.6	356	352
0. orca	3,006	6,642	45.3	553	566
S. attenuata	353	711	49.6		304^{a}
S. longirostris	247	541	45.6	_	286ª
S. bredanensis	706	1,454	48.6	378	388
T. truncatus	685	1,550	44.2	376	377
Monodontidae					
D. leucas	938	2,087	44.9	456	414
Phocoenidae					
P. phocoena	242	506	47.7	316	266
P. dalli	270	803	33.6	334	282
Physeteridae					
P. macrocephalus	3,308	7,693	43	547	582
Pontoporiidae					
P. blainvillei	154.9	223.9	69.2	319	271
Ziphiidae					
M. europaeus	971	1,680	57.8		
Balaenopteridae					
B. physalus	2,640	6,718	39.3	342	537
Bovidae					
B. taurus	199 ^b	456 ^b	43.6	278°	270
O. aries	69	130 ^d	53	150 ^e	208
Giraffidae					
G. camelopardalis	428 ^f	537 ^f	79.7	459 ^c	363
Hippopotamidae					
H. amphibius	195 ^ь	590 ^b	33.1	240 ^e	258
Hominidae					
H. sapiens	380 ^g	$1,400^{b}$	27	280 ^e	324

^aPerrin *et al.* (1977).

^bSacher and Staffeldt (1974).

^cKiltie (1982).

^dMinervini et al. (2016).

eHayssen et al. (1993).

Graic et al. (2017).

^gBlinkov and Glezer (1968).



Figure 5. One example each of a male neonate brain of 0. orca compared with a brain of a male adult. The ratio of cerebellum mass to the mass of the whole brain is similar in the neonate and adult. The mass of the neonate brain is 3,292 g, while the adult brain mass is 7,100 g.

periods, neonatal and adult cetaceans have much larger brains than their terrestrial counterparts (see Table 2).

For example, the neonatal elephant brain may be over 1,600 g (Pagel and Harvey 1988). Shoshani *et al.* (2006) gave one value of 1,724 g for a neonatal elephant brain. Our neonatal killer whales had brains almost twice as large as the neonatal elephant. The elephant gestation, at around 656 d (Wittemyer *et al.* 2007), is even longer than that of the killer whale, at 553 d. The neonatal killer whale brain grows to a mass that is almost twice as large as the neonatal elephant's brain within a much shorter period of gestation.

Underwater Acoustics: Hearing and Sonar

Members of the family Delphinidae have the largest relative total brain size among cetaceans. They also have the largest relative cerebellum size (Ridgway et al. 2017). Like other cetaceans, delphinids live their entire lives in the water where sound travels over four times as fast as in air at the same temperature. This simple fact impacts every aspect of cetacean existence, including brain development. According to Langworthy's (1932) study of the *T. truncatus* brain, numerous tracts of acoustic fibers reach the cerebral cortex, allowing the cortex to reach an advanced stage of development on the basis of these acoustic impulses.

Bats also use sonar. Since they do not have large brains some have suggested that the bat's brain size is an argument against the involvement of acoustic capability in the explanation for the large brain size of delphinids (e.g., Manger 2006, Marino 2007). However, there are many differences in using sonar (echolocation) between bats in air and cetaceans in water. Worthy and Hickie (1986) summarized the findings of Wood and Evans (1980) and suggested various differences between bat and delphinid echolocation. These included differences in sonar sound bandwidth (tonal sounds vs. broadband clicks). Delphinids need to detect targets such as fish and squid

that are very similar in density to their water environment. The intervals between delphinid sonar click emissions are very short. Short intervals for reception of echoes from closely spaced but separate objects require rapid processing by the brain. Other comparisons have been made based on brain size and neural responses (Ridgway 1986, Huggenberger 2008).

Perhaps an even greater difference between bat and dolphin brain expansion has to do with the acoustic environment during development. The cetacean body, except for the lungs and other air-containing spaces, is a close impedance match to water (Au 1996). It is highly likely that sound from the environment passes into the pregnant cetacean's body through the amniotic fluid to the developing cetacean. This may drive the early development of the auditory system. The dolphin brain's auditory fibers are already myelinated in the womb long before the cetacean is born (Hosokawa et al. 1969, Solntseva 1999, Montie et al. 2007). Early myelination of the auditory system suggests that the cetacean fetus can hear. If a sound source is coupled to the mother's body, the human fetus can hear at a gestational age of about 29 wk (Moore et al. 1995). At this time, the auditory nervous system is at least partially myelinated. It is likely that the dolphin fetus can hear sounds from the ocean environment as its mother swims.

Audiograms of the developing cetacean fetus have not yet been done. Many ultrasound examinations of developing dolphins are done each year so, using methods analogous to those used by Moore *et al.* (1995), fetal audiograms should be possible. Coupling of sounds from the water may allow the fetus to hear its mother and nearby animals within the group. The developing cetacean inside the womb may hear a wide range of sounds long before it is born (Mello and Amundin 2005). This could be an advantage for cetaceans that live in groups compared to solitary species. Fetal sound reception very possibly drives the extreme development of the cetacean auditory nervous system, empowering cetaceans to maximize their use of sound within the marine environment. Thus, fetal hearing tests might link delphinids' early auditory ability and the development of a large brain.

Metabolism, Energetics, and Life Span

Our data show that neonatal brain mass scales to between 0.5 and 0.6 power of maternal body mass (Fig. 1). Similarly, delphinid cortex surface area scales to about the 0.6 power of body mass (Ridgway *et al.* 2017). In contrast, Martin (1981) found that neonatal brain mass in terrestrial mammals scales to the 0.75 power of maternal body mass. The terrestrial's low calorie food compared to the delphinid mother's high calorie food is a major difference between the groups. Large amounts of high calorie food facilitate development of large brained delphinid offspring and may help to support brain growth during a long period of nursing.

Hofman (1993) hypothesized that encephalization and metabolism may direct the evolutionary course of lifespan, growth and reproduction. Hofman's hypothesis is supported by the elephant's high encephalization, long life span, and its relatively low metabolic rate. Elephants consume large amounts of plant material, but their food is relatively low in caloric value. Christiansen (2004) states "both the basal and the field metabolic rates of extant elephants are lower than predicted for a hypothetical mammal, in accordance with their body size and subsistence on low-quality foods." In contrast, the killer whale, the largest delphinid, does not appear to support Hofman's (1993) hypothesis. Killer whales and elephants are both highly encephalized, but killer whales consume many more calories each day (Noren 2011, Williams

et al. 2017). Neonatal cetacean brains, such as those of sperm and killer whales, grow to twice the size of the neonatal elephant brain within a shorter period of gestation.

Measurements of oxygen and glucose consumption revealed that delphinids have a higher metabolic rate compared to most terrestrial mammals, and measures of the aerobic costs of swimming and diving demonstrate the delphinids' high energy lifestyle (Ridgway 1990, Williams et al. 2001, Ridgway et al. 2006, Noren 2011, Williams et al. 2017). The metabolic rate of adult bottlenose dolphins and killer whales is much higher than the value predicted by Kleiber's law, which states that an animal's metabolic rate scales to the 0.75 power of its body mass (Noren 2011, Williams et al. 2017). A delphinid's high energy aquatic lifestyle and its abilities to swim fast and chase elusive prey underwater are enabled by a high-calorie diet that is digested by long alimentary tracts (Williams et al. 2001). We suggest that the rich, high calorie food supply of cetaceans has enabled them to support the metabolic demands of a large brain.

Conclusion

We found that delphinid brains grow to between 1.5 and 10 times as large as similar sized terrestrial cetartiodactyls. Delphinids' larger brain growth during life at sea may be explained by at least three differences from terrestrial cetartiodactyls' life on land. First, the sea offers high calorie prey to support growth of a large brain. Second, life in water offers relief from gravity, allowing for a large head to contain a large brain. Third, sound in water may pass through an immersed body. This allows sounds from the water to reach the fetus, driving early development of delphinoid auditory brains to enable development of an effective sonar system.

The data presented herein expand upon the data that are currently available in the literature on cetacean brain size and growth across maturity and body size. In our data set, we included measures of brain mass, brain volume, body mass, gestation duration, body length, and EQ. We found a positive and direct correlation between gestation duration and brain volume for neonates of Delphinidae, the taxonomic family from which the majority of our data came. Neonatal brain mass is also positively and directly correlated with maternal body mass in delphinids. Upon investigating the trajectory of brain growth from birth to maturity, we found that EQ decreases as body length increases and brain mass appears to grow beyond sexual maturity in most of the odontocete species we examined. There is a slower rate of brain growth from neonatal to adult length for larger delphinids, such as *Orcinus orca*, than for smaller delphinids and members of the family Phocoenidae. The expanded data set of the present study offers a platform for future investigations on cetacean brain development across the lifespan.

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LITERATURE CITED

- Atkinson, S., and M. Yoshioka. 2007. Endocrinology of reproduction. Pages 171–192 in D. L. Miller, ed. Reproductive biology and phylogeny of Cetacea. Science Publishers, Enfield, NH.
- Au, W. W. 1996. Acoustic reflectivity of a dolphin. Journal of the Acoustical Society of America 99:3844–3848.
- Ballarin, C., M. Povinelli, A. Granato, M. Panin, L. Corain, A. Peruffo and B. Cozzi. 2016. The brain of the domestic *Bos taurus*: Weight, encephalization and cerebellar quotients, and comparison with other domestic and wild Cetartiodactyla. PLOS ONE 11: e0154580.
- Barton, R. A., and I. Capellini. 2011. Maternal investment, life histories and the costs of brain growth in mammals. Proceedings of the National Academy of Sciences of the United States of America 108:6169–6174.
- Berta, A., J. L. Sumich and K. M. Kovacs. 2005. Marine mammals: Evolutionary biology. Academic Press, New York, NY.
- Blinkov, S. M., and I. I. Glezer. 1968. The human brain in figures and tables. Plenum Press, New York, NY.
- Boddy, A. M., M. R. McGowen, C. C. Sherwood, L. I. Grossman, M. Goodman and D. E. Wildman. 2012. Comparative analysis of encephalization in mammals reveals relaxed constraints on anthropoid primate and cetacean brain scaling. Journal of Evolutionary Biology 25:981–994.
- Brodie, P. F. 1969. Duration of lactation in Cetacea: An indicator of required learning? The American Midland Naturalist 82:312–314.
- Caldwell, D. K., and M. C. Caldwell. 1989. Pygmy sperm whale *Kogia breviceps* (de Blainville, 1838): Dwarf sperm whale *Kogia simus* Owen, 1866. Pages 235–260 in S. H. Ridgway and R. Harrison, eds. Handbook of marine mammals. Volume 4. River dolphins and the larger toothed whales. Academic Press, London, U.K.
- Charvet, C. J., and B. L. Finlay. 2012. Embracing covariation in brain evolution: Large brains, extended development, and flexible primate social systems. Progress in Brain Research 195:71–87.
- Christiansen, P. 2004. Body size in proboscideans, with notes on elephant metabolism. Zoological Journal of the Linnean Society 140:523–549.
- Connor, R. C. 2007. Dolphin social intelligence: Complex alliance relationships in bottlenose dolphins and a consideration of selective environments for extreme brain size evolution in mammals. Philosophical Transactions of the Royal Society of London, Series B: Biological Sciences 362:587–602.
- Dahlheim, M. E., and J. E. Heyning. 1999. Killer whale *Orcinus orca* (Linnaeus, 1758). Pages 281–322 *in* S. H. Ridgway and R. Harrison, eds. Handbook of marine mammals.

- Volume 6. The second book of dolphins and the porpoises. Academic Press, London, U.K.
- DeGraaf, A. S. 1967. Anatomical aspects of the cetacean brain stem. Van Gorcum and Company, Assen, The Netherlands.
- Dekaban, A. S., S. Anatole and D. Sadowsky. 1978. Changes in brain weights during the span of human life: Relation of brain weights to body heights and body weights. Annals of Neurology 4:345–356.
- DeSilva, J., and J. Lesnik. 2006. Chimpanzee neonatal brain size: Implications for brain growth in *Homo erectus*. Journal of Human Evolution 51:207–212.
- Dobbing, J., and J. Sands. 1979. Comparative aspects of the brain growth spurt. Early Human Development 311:79–83.
- George, J. C., J. Bada, J. Zeh, L. Scott, S. E. Brown, T. O'Hara and R. Suydam. 1999. Age and growth estimates of bowhead whales (*Balaena mysticetus*) via aspartic acid racemization. Canadian Journal of Zoology 77:571–580.
- Gingerich, P. D., M. ul Haq, I. S. Zalmout, I. H. Khan and M. S. Malkani. 2001. Origin of whales from early artiodactyls: Hands and feet of Eocene Protocetidae from Pakistan. Science 293:2239–2242.
- Gompertz, R. H. C. 1902. Specific gravity of the brain. The Journal of Physiology 27:459–462.
- Graïc, J. M., A. Peruffo, C. Ballarin and B. Cozzi. 2017. The brain of the giraffe (*Giraffa camelopardalis*). Surface configuration, encephalization quotient, and analysis of the existing literature. The Anatomical Record 300:1502–1511.
- Hanson, A., W. Grisham, C. Sheh, J. Annese and S. Ridgway. 2013. Quantitative examination of the bottlenose dolphin cerebellum. The Anatomical Record 296:1215–1228.
- Harvey, P. H., and J. R. Krebs. 1990. Comparing brains. Science 249:140-146.
- Hayssen, V., A. van Tienhoven and A. van Tienhoven. 1993. Asdell's patterns of mammalian reproduction: A compendium of species-specific data. Cornell University Press, Ithaca, NY
- Herculano-Houzel, S., K. Catania, P. R. Manger and J. H. Kaas. 2015. Mammalian brains are made of these: A dataset of the numbers and densities of neuronal and nonneuronal cells in the brain of glires, primates, scandentia, eulipotyphlans, afrotherians and artiodactyls, and their relationship with body mass. Brain, Behavior and Evolution 86:145–163.
- Hofman, M. A. 1993. Encephalization and the evolution of longevity in mammals. Journal of Evolutionary Biology 6:209–227.
- Hosokawa, H., S. Igarashi, T. Kamija and K. Hirosawa. 1969. Morphological characteristics and myelinization of acoustic system in the dolphins (*Stenella coeruleoalba*). Scientific Reports of the Whales Research Institute, Tokyo 21:95–123.
- Houck, W. J., and T. A. Jefferson. 1999. Dall's porpoise *Phocoenoides dalli* (True, 1885). Pages 443–472 in S. H. Ridgway and R. Harrison, eds. Handbook of marine mammals. Volume 6. The second book of dolphins and the porpoises. Academic Press, London, U.K.
- Huggenberger, S. 2008. The size and complexity of dolphin brains—a paradox? Journal of the Marine Biological Association of the United Kingdom 88:1103–1108.
- Isler, K., and C. P. van Schaik. 2012. Allomaternal care, life history and brain size evolution in mammals. Journal of Human Evolution 63:52–63.
- Jerison, H. J. 1973. Evolution of the brain and intelligence. Academic Press, New York, NY.
- Kiltie, R. A. 1982. Intraspecific variation in the mammalian gestation period. Journal of Mammalogy 63:646–652.
- Lacave, G., M. Eggermont, T. Verslycke, F. Brook, A. Salbany, L. Roque and R. Kinoshita. 2004. Prediction from ultrasonographic measurements of the expected delivery date in two species of bottlenosed dolphin (*Tursiops truncatus* and *Tursiops aduncus*). Veterinary Record 154:228–233.

- Langworthy, O. R. 1932. A description of the central nervous system of the porpoise (*Tursiops truncatus*). Journal of Comparative Neurology 52:437–498.
- Laws, R. M. 1959. The foetal growth rates of whales with special reference to the fin whale, *Balaenoptera physalus* Linn. Discovery Reports 29:281–408.
- Lockyer, C. 2007. All creatures great and smaller: A study in cetacean life history energetics. Journal of the Marine Biological Association of the United Kingdom 87:1035–1045.
- Mallette, S. D., W. A. McLellan, F. S. Scharf, H. N. Koopman, S. G. Barco, R. S. Wells and D. A. Pabst. 2016. Ontogenetic allometry and body composition of the common bottlenose dolphin (*Tursiops truncatus*) from the U. S. mid-Atlantic. Marine Mammal Science 32:86–121.
- Manger, P. R. 2006. An examination of cetacean brain structure with a novel hypothesis correlating thermogenesis to the evolution of a big brain. Biological Reviews 81:293–338.
- Marino, L. 1997. The relationship between gestation length, encephalization, and body weight in odontocetes. Marine Mammal Science 13:133–138.
- Marino, L. 2007. Cetacean brains: How aquatic are they? The Anatomical Record 290:694–700.
- Marino, L., J. K. Rilling, S. K. Lin and S. H. Ridgway. 2000. Relative volume of the cerebellum in dolphins and comparison with anthropoid primates. Brain, Behavior and Evolution 56:204–211.
- Marino, L., D. W. McShea and M. D. Uhen. 2004. The origin and evolution of large brains in toothed whales. The Anatomical Record 281A:1247–1255.
- Marino, L., M. D. Uhen, B. Frohlich, J. M. Aldag, C. Blane, D. Bohaska and F. C. Whitmore, Jr. 2000. Endocranial volume of mid-late Eocene Archaeocetes (Order: Cetacea) revealed by computed tomography: Implications for cetacean brain evolution. Journal of Mammalian Evolution 7:81–94.
- Martin, R. D. 1981. Relative brain size and basal metabolic rate in terrestrial vertebrates. Nature 293:57–60.
- McLellan, W. A., H. N. Koopman, S. A. Rommel, et al. 2002. Ontogenetic allometry and body composition of harbour porpoises (*Phocoena phocoena*, L.) from the western North Atlantic. Journal of Zoology 257:457–471.
- Mello, L., and M. Amundin. 2005. Whistle production pre- and post-partum in bottlenose dolphins (*Tursiops truncatus*) in human care. Aquatic Mammals 31:169–175.
- Minervini, S., G. Accogli, A. Pirone, J. M. Graïc, B. Cozzi and S. Desantis. 2016. Brain mass and encephalization quotients in the domestic industrial pig (Sus scrofa). PLOS ONE 11:1–17.
- Miyazaki, N., Y. Fujise and T. Fujiyama. 1981. Body and organ weight of striped and spotted dolphins off the Pacific coast of Japan. Scientific Reports of the Whales Research Institute, Tokyo 33:27–67.
- Montgomery, S. H., J. H. Geisler, M. R. McGowen, C. Fox, L. Marino and J. Gatesy. 2013. The evolutionary history of cetacean brain and body size. Evolution 67:3339–3353.
- Montie, E. W., G. Schneider, D. R. Ketten, L. Marino, K. E. Touhey and M. E. Hahn. 2007. Neuroanatomy of the subadult and fetal brain of the Atlantic white-sided dolphin (*Lagenorhynchus acutus*) from in situ magnetic resonance images. The Anatomical Record 290:1459–1479.
- Moore, J. K., L. M. Perazzo and A. Braun. 1995. Time course of axonal myelination in the human brainstem auditory pathway. Hearing Research 91:208–209.
- Noren, D. P. 2011. Estimated field metabolic rates and prey requirements of resident killer whales. Marine Mammal Science 27:60–77.
- O'Brien, J. K., and T. R. Robeck. 2012. The relationship of maternal characteristics and circulating progesterone concentrations with reproductive outcome in the bottlenose dolphin (*Tursiops truncatus*) after artificial insemination, with and without ovulation induction, and natural breeding. Theriogenology 78:469–482.

- Pagel, M., and P. Harvey. 1988. How mammals produce large-brained offspring. Evolution 42:948–957.
- Passingham, R. 1975. Changes in the size and organisation of the brain in man and his ancestors. Brain, Behavior and Evolution 11:73–90.
- Perrin, W. F., and J. W. Gilpatrick. 1994. Spinner dolphin Stenella longirostris (Gray, 1828).
 Pages 99–128 in S. H. Ridgway and R. Harrison, eds. Handbook of marine mammals.
 Volume 5. The first book of dolphins. Academic Press, London, U.K.
- Perrin, W. F., D. B. Holts and R. B. Miller. 1977. Growth and reproduction of the eastern spinner dolphin, a geographical form of *Stenella longirostris* in the eastern tropical Pacific. Fishery Bulletin 75:725–750.
- Perrin, W. F., R. L. Brownell and D. P. DeMaster. 1984. Reproduction in whales, dolphins and porpoises. Report of the International Whaling Commission (Special Issue 6).
- Perrin, W. F., C. E. Wilson and F. I. Archer. 1994. Striped dolphin *Stenella coeruleoalba* (Meyen, 1833). Pages 129–159 *in* S. H. Ridgway and R. Harrison, eds. Handbook of marine mammals. Volume 5. The first book of dolphins. Academic Press, London, U.K.
- Reese, C. S., J. A. Calvin, J. C. George and R. J. Tarpley. 2001. Estimation of fetal growth and gestation in bowhead whales. Journal of the Acoustical Society of America 96:906–914
- Ridgway, S. H. 1981. Some brain morphometrics of the bowhead whale. Pages 837–844 in T. F. Albert, ed. Tissue structural studies and other investigations on the biology of endangered whales in the Beaufort Sea. Volume 1. Final report to the Bureau of Land Management, U.S. Department of Interior. University of Maryland, College Park, MD.
- Ridgway, S. H. 1986. Dolphin cognition and behavior: a comparative approach. Pages 31–60 in R. J. Schusterman, J. A. Thomas and F. G. Wood, eds. Physiological observations on dolphin brains. Lawrence Erlbaum, Hillsdale, NJ.
- Ridgway, S. H. 1990. The central nervous system of the bottlenose dolphin. Pages 69–97 in S. Leatherwood and R. Reeves, eds. The bottlenose dolphin, *Tursiops* spp. Academic Press, New York, NY.
- Ridgway, S. H., and A. C. Hanson. 2014. Sperm whales and killer whales with the largest brains of all toothed whales show extreme differences in cerebellum. Brain, Behavior and Evolution 83:266–274.
- Ridgway, S. H., and R. J. Tarpley. 1996. Brain mass comparisons in Cetacea. Conference Proceedings of the International Association of Aquatic Animal Medicine 27:55–57.
- Ridgway, S. H., and K. R. Van Alstyne. 2017. The blue whale brain misrepresented by an alcohol dehydrated brain of 3,636 grams. Marine Mammal Science 33:386–388.
- Ridgway, S. H., H. J. Flanigan and J. G. McCormick. 1966. Brain-spinal cord ratios in porpoises: Possible correlations with intelligence and ecology. Psychonomic Science 6:491–492.
- Ridgway, S. H., D. Houser, J. J. Finneran, et al. 2006. Functional imaging of dolphin brain metabolism and blood flow. Journal of Experimental Biology 209:2902–2910.
- Ridgway, S. H., K. P. Carlin, K. R. Van Alstyne, A. C. Hanson and R. J. Tarpley. 2017. Comparison of dolphins' body and brain measurements with four other groups of cetaceans reveals great diversity. Brain, Behavior and Evolution 88:235–257.
- Robeck, T. R., K. J. Steinman, S. Gearhart, T. R. Reidarson, J. F. McBain and S. L. Monfort. 2004. Reproductive physiology and development of artificial insemination technology in killer whales (*Orcinus orca*). Biology of Reproduction 71:650–660.
- Robeck, T. R., K. J. Steinman, M. Greenwell, et al. 2009. Seasonality, estrous cycle characterization, estrus synchronization, semen cryopreservation, and artificial insemination in the Pacific white-sided dolphin (Lagenorhynchus obliquidens). Reproduction 138:391–405.
- Sacher, G. A., and E. F. Staffeldt. 1974. Relation of gestation time to brain weight for placental mammals: Implications for the theory of vertebrate growth. The American Naturalist 108:593–615.

- Shoshani, J., W. J. Kupsky and G. H. Marchant. 2006. Elephant brain: Gross morphology, functions, comparative anatomy, and evolution. Brain Research Bulletin 70:124–157.
- Silva, M., and J. A. Downing. 1995. The allometric scaling of density and body mass: A nonlinear relationship for terrestrial mammals. American Naturalist 145:704–727.
- Solntseva, G. 1999. The comparison of the development of the auditory and vestibular structures in toothed whales—beluga (*Delphinapterus leucas*, Cetacea: Odontoceti). Doklady Akademii Nauk 364:714–718.
- Tarpley, R., and S. H. Ridgway. 1994. Corpus callosum size in delphinid cetaceans. Brain, Behavior and Evolution 44:156–165.
- Weisbecker, V., and A. Goswami. 2010. Neonatal maturity as the key to understanding brain size evolution in homeothermic vertebrates. Bioessays 33:155–158.
- Wells, R. S., and M. D. Scott. 1999. Bottlenose dolphin *Tursiops truncatus* (Montagu, 1821). Pages 137–182 *in* S. H. Ridgway and R. Harrison, eds. Handbook of marine mammals. Volume 6. The second book of dolphins and the porpoises. Academic Press, London, U.K.
- Williams, T. 2006. Physiological and ecological consequences of extreme body size in whales. Pages 306–309 *in* J. A. Estes, ed. Whales, whaling, and ocean ecosystems. University of California Press, Berkeley, CA.
- Williams, T. M., J. Haun, R. W. Davis, L. A. Fuiman and S. Kohin. 2001. A killer appetite: Metabolic consequences of carnivory in marine mammals. Comparative Biochemistry and Physiology Part A: Molecular and Integrative Physiology 129:785–796.
- Williams, T. M., T. L. Kendall, B. P. Richter, *et al.* 2017. Swimming and diving energetics in dolphins: A stroke-by-stroke analysis for predicting the cost of flight responses in wild odontocetes. Journal of Experimental Biology 220:1135–1145.
- Wittemyer, G., H. B. Rasmussen and I. Douglas-Hamilton. 2007. Breeding phenology in relation to NDVI variability in free-ranging African elephant. Ecography 30:42–50.
- Wood, F. G., and W. E. Evans. 1980. Adaptiveness and ecology of echolocation in toothed whales. Pages 381–425 *in* R. G. Busnel and J. F. Fish, eds. Animal sonar systems. Springer, New York, NY.
- Worthy, G. A. J., and J. P. Hickie. 1986. Relative brain size in marine mammals. The American Naturalist 128:445–459.
- Xie, Z. H., S. G. Sun, Q. S. She, L. Y. Chen and J. Wang. 2011. Stereological estimation of volumes and cortical surface areas of the cerebrum and cerebellum in fixed bactrian camel (*Camelus bactrianus*) brain. Pages 355–360 in T. Gaholt, A. Saber, S. Nagpal and J. Wang, eds. Selected research on gross anatomy and histology of camels. Camel Publishing House, Bikaner, India.

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SUPPORTING INFORMATION

The following supporting information is available for this article online at http://onlinelibrary.wiley.com/doi/10.1111/mms.12464/suppinfo.

Table S1. Data set of all cetaceans used in our analyses. Sexes, ages (adult, subadult, or neonate), body lengths, brain and body masses, gestation durations, taxonomic classifications, and origin of data are given for individuals of 52 cetacean species (9 mysticete, 43 odontocete). Counts of specimens per species from each of three age groups (adult, subadult, and neonate) and from each of three sources of data origin (author, literature, and other) are given under the "Brain data-species counts" tab. A separate table comparing encephalization indices for neonate and adult female cetaceans (and humans) is included under the "Neonate-Adult Brain-Body mass" tab.